

Replacement of a Thiourea-S with an Amidine-NH Donor Group in a Platinum–Acridine Antitumor Compound Reduces the Metal's Reactivity with Cysteine Sulfur

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The reactivity of two DNA-targeted platinum–acridine conjugates with cysteine sulfur was studied. The conjugate containing an amidine-NH donor group cis to the chloride leaving group showed considerably reduced reactivity with *N*-acetylcysteine compared to the prototypical derivative containing a thiourea-S linkage. The opposite scenario has been observed previously in reactions with nucleobase nitrogen. Possible consequences of the unique target-selective tuning of the substitution chemistry for the pharmacodynamic properties and biological activity of these agents are discussed.

Introduction

The high reactivity of the anticancer drug cisplatin (*cis*-[PtCl₂(NH₃)₂]) with the intracellular sulfur-containing nucleophile glutathione (γ -L-glutamyl-L-cysteinylglycine, GSH⁴) causes adverse effects such as severe systemic toxicities and tumor resistance.^{1,2} One major goal in platinum drug development, therefore, is to design complex geometries that react less avidly with cysteine sulfur. This has been achieved, for instance, in picoplatin (*cis*-[PtCl₂(NH₃)(2-methylpyridine)]) by replacing one ammine ligand in cisplatin with a sterically hindered pyridine derivative,^{3,4} and in carboplatin (*cis*-[Pt(CBDCA)(NH₃)₂], CBDCA = cyclobutane-1,1-dicarboxylate) by introducing a bidentate leaving group in place of the chlorido ligands. While carboplatin is remarkably inert in reactions with GSH,⁵ the stability of the dicarboxylate chelate compromises the metal's ability to bind to its pharmacological target, DNA.⁶ This dilemma is manifest in the fact that the second-generation drug, while less toxic than cisplatin, has to be administered at significantly higher doses to achieve the same therapeutic effect as the parent drug.⁷

Here we report a unique case of a chemical modification to a DNA-targeted platinum agent that *reduces* the reactivity of the metal with cysteine sulfur while it was previously found to *accelerate* the reaction with nucleobase nitrogen. In a recent study,⁸ we investigated the biological effect of replacing a thiourea (*sp*²-S) with an amidine (*sp*²-NH) donor cis to a chloride leaving group. The new platinum-acridine agent reported, [PtCl(en)(L)](NO₃)₂ (en = ethane-1,2-diamine; L = *N*-(2-(acridin-9-ylamino)ethyl)-*N*-methylpropionamidinium, acridinium cation) (compound **1**, Figure 1) proved to be a significantly more efficient DNA binder and more cytotoxic agent than the prototypical agent, PT-ACRAMTU^{9,10} (ACRAMTU = 1-(2-(acridin-9-ylamino)ethyl)-1,3-dimethylthiourea, acridinium cation) (compound **2**, Figure 1). Moreover, this work has led to the first member of this class of compounds endowed with antitumor activity in aggressive non-small cell lung cancer in a mouse xenograft model.⁸ To test the effect of this simple structural modification on the metal's reactivity with protein sulfur, we incubated both analogues with *N*-acetylcysteine (*N*-

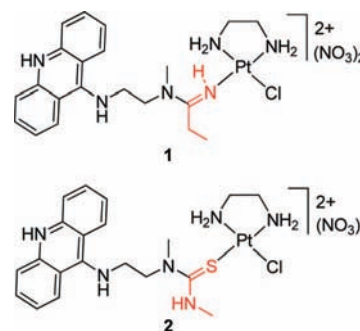


Figure 1. Structures of compound **1** and PT-ACRAMTU (**2**) with structural differences highlighted.

AcCys), a modified amino acid mimicking GSH, and monitored the progress of the reactions by ¹H NMR spectroscopy. In addition, we analyzed the mixtures after completion of the reactions by in-line high-performance liquid chromatography electrospray mass spectrometry (LC-MS).

Results and Discussion

In arrayed one-dimensional ¹H NMR experiments, the reaction of complex **1** with 1 equiv of *N*-AcCys (25 °C, 10 mM phosphate buffer, D₂O, pH* 6.8) was found to proceed considerably more slowly than the reaction of PT-ACRAMTU (**2**) under the same conditions. The relative rates at which the amino acid reacted with **1** and **2** were deduced from the change in integral intensities of ¹H NMR signals assigned to the platinum complexes (see Supporting Information for spectral data and detailed procedures). Under the conditions chosen, the reaction between **1** and *N*-AcCys follows a second-order rate law, typical of a mechanism that is dominated by direct substitution of the chlorido ligand by cysteine sulfur and is not limited by the rate of aquation.² In contrast, the more rapid reaction of PT-ACRAMTU (**2**) with *N*-AcCys followed neither second-order nor first-order kinetics, suggesting that the reaction of this derivative proceeds via a more complicated mechanism (Figure 2). In both cases, the ¹H NMR spectra of the mixtures indicated formation of multiple products. Attempts to assign the species formed by **1** and **2** using two-dimensional [¹H–¹⁵N] NMR spectroscopy, a technique applied previously in nucleotide binding studies of [¹⁵N]-en labeled conjugates,^{8,11} were unsuccessful due to the complexity of the reaction mixtures.

To overcome this difficulty, LC-MS was used to analyze the product distribution in mixtures of platinum complex incubated

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^a Abbreviations: *N*-AcCys, *N*-acetylcysteine; ACRAMTU, 1-(2-(acridin-9-ylamino)ethyl)-1,3-dimethylthiourea; GSH, glutathione; pH*, uncorrected pH meter reading in D₂O.

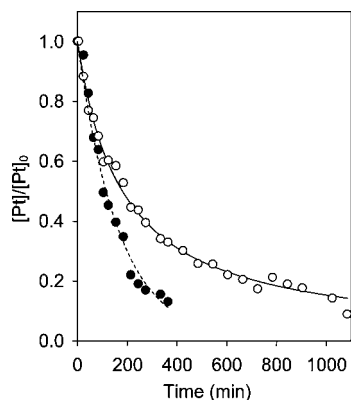


Figure 2. Progress of the reactions of compound **1** (open circles) and compound **2** (filled circles) with 1 equiv of *N*-acetylcysteine (*N*-AcCys) at 25 °C monitored by ^1H NMR spectroscopy. The data shown is the mean of two experiments. The solid line (for **1**) represents a curve fit assuming second-order conditions, $[\text{Pt}]/[\text{Pt}]_0 = (1 + k[\text{Pt}]_0 t)^{-1}$; ($t_{1/2} = 180$ min). The dashed trend line (for **2**) has no physical significance.

with 1 equiv of sulfur nucleophile (37 °C, 6 h, 10 mM phosphate buffer, pH 7.2). The two most abundant adducts formed by compound **1** were unambiguously identified as the mononuclear species $[\text{Pt}(\text{en})(\text{L})(\text{N-AcCys}^*)]^+$ (**I**) $^+$, 14%) and the dinuclear

complex $[\{\text{Pt}(\text{en})(\text{L})\}_2(\mu\text{-N-AcCys}^*)]^{4+}$ (**II**) $^{4+}$, 72%) in electrospray mass spectra recorded in positive-ion mode (Figure 3). (The asterisk indicates the dianionic form of metal-bound *N*-AcCys at neutral pH.) These findings suggest that adduct **I**, formed by substitution of chloride by cysteine sulfur, rapidly reacts with unreacted complex **1** to form the dinuclear species, **II** (Scheme 1). The tendency of cysteine sulfur to readily induce bridged complexes with platinum(II) drugs has been amply demonstrated.² The platinum–amidine linkage and the en chelate in complex **1** appear to be resistant to nucleophilic attack by cysteine based on the presence of only a minor amount of free acridine **L** (**III**, 3%) and the absence of ring-opened adducts. In contrast, the LC-MS data acquired for PT-ACRAMTU (**2**) reveal a more complicated reaction pattern. In analogy to complex **1**, the chloride substitution pathway leads to the dinuclear adduct, $[\{\text{Pt}(\text{en})(\text{ACRAMTU})\}_2(\mu\text{-N-AcCys}^*)]^{4+}$ (**II'**, 32%), most likely via intermediate **I'**, which could not be detected in this case (Figure 3). In addition, significant amounts of ACRAMTU (**III'**, 33%) and the bisintercalator complex, $[\text{Pt}(\text{en})(\text{ACRAMTU})_2]^{4+}$ (**IV**, 26%) (Scheme 1), were observed in the HPLC profiles (Supporting Information). Formation of the latter complex can be explained with chloride substitution in compound **2** by free ACRAMTU in the reaction mixture. The release of ACRAMTU by *N*-AcCys was unexpected and

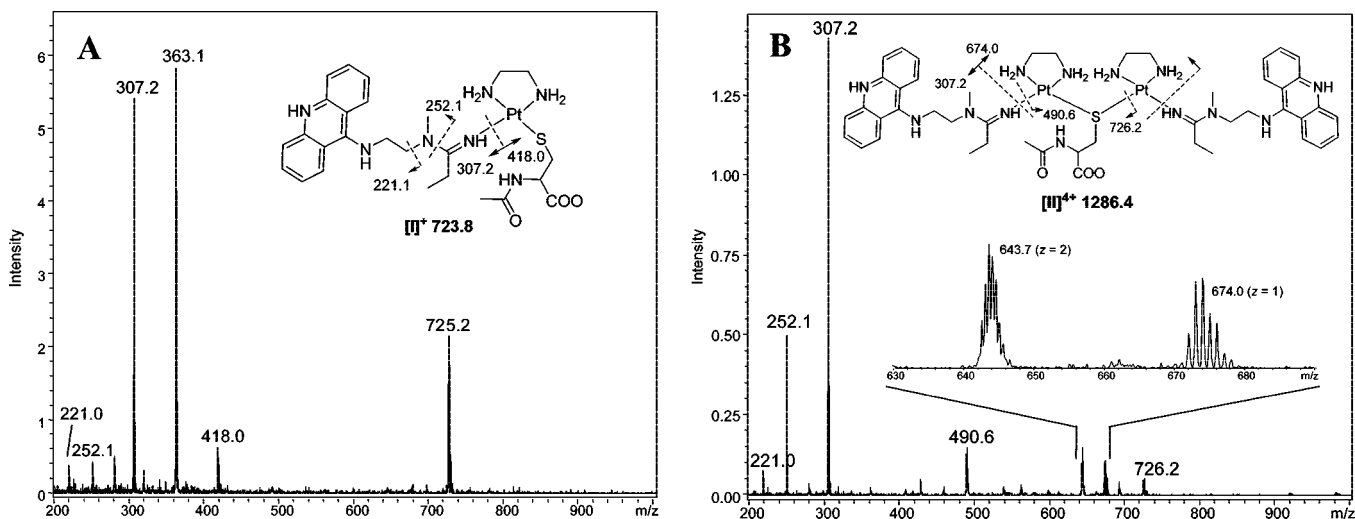
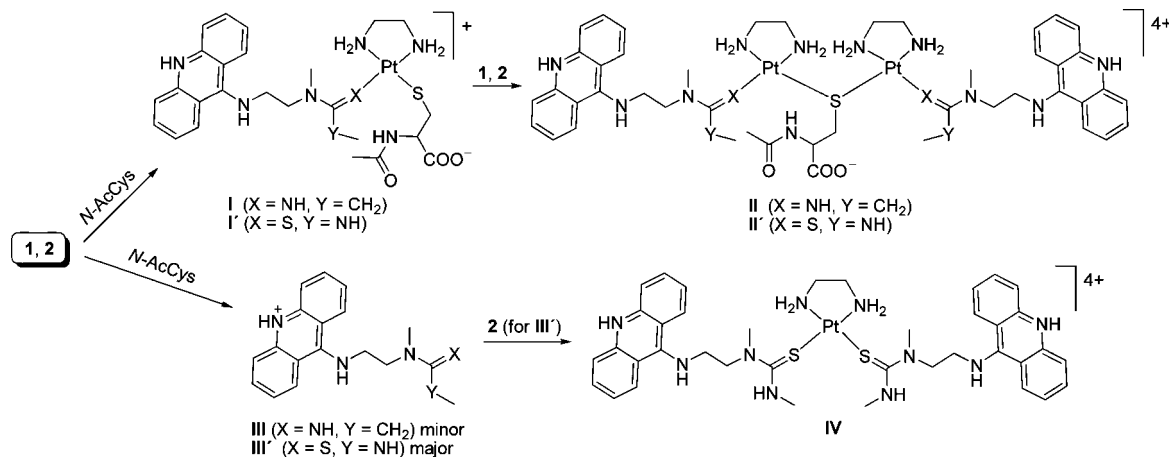


Figure 3. Positive-ion electrospray mass spectra of adducts **I** (A) and **II** (B) showing peaks for the molecular ions, **I** $^+$ (m/z 725.2) and **II**- 2H $^{2+}$ (m/z 643.7, $z = 2$) and fragment ions resulting from in-source collisionally activated dissociation (CAD). The insets illustrate the observed fragmentation patterns. The discrepancy between the calculated and observed m/z values for some of the platinum containing ions is due to distorted Pt isotope patterns.

Scheme 1. Reactivity of Compounds **1** and **2** with *N*-Acetylcysteine (*N*-AcCys)



shows that thiourea, while a typical nonleaving group in reactions of compound **2** with DNA nitrogen,^{12,13} becomes substitution-labile in the presence of cysteine sulfur.

The reactivity of the Pt–S_{thiourea} bond and its cis labilizing effect on chloride are potential contributors to the increased cysteine reactivity of PT-ACRAMTU (**2**) compared to analogue **1**. The rationale for incorporating a thiourea-based nonleaving group in the original design was to enhance the reactivity of the metal with DNA nucleobases by exploiting the cis activating effect of sulfur. In their pioneering studies of the inorganic kinetic cis-effect, Tobe and co-workers demonstrated for the complexes [PtCl(en)(dms_o-S)]⁺ and [PtCl(en)(dms)]⁺ (dms_o = dimethylsulfoxide, dms = dimethylsulfide) that the sulfur donors greatly accelerate chloride substitution by an incoming nucleophile relative to analogous complexes with [N₃Cl] donor sets.^{14,15} In the same studies, the authors demonstrated that the relative rate enhancement (k_S/k_N) is large for strong incoming nucleophiles, such as sulfur ligands, but negligible for weak ligands like water. The comparative kinetic study of amidine-modified complex **1** and thiourea-based PT-ACRAMTU (**2**) shows that replacement of the sulfur with an imino donor group reduces, as expected, the metal's reactivity with cysteine sulfur but enhances its binding with DNA nitrogen. These findings strongly suggest that, unlike the reaction with cysteine, the reaction with DNA is not controlled by the electronic cis-effect but, as concluded previously, most likely by steric factors and/or H-bonding that favor complex aquation and facilitate nucleophilic attack of nucleobase nitrogen on the metal center.⁸

In addition to the unique monofunctional-intercalative adducts formed by PT-ACRAMTU and its second-generation analogues, one critical feature that sets these hybrid agents apart from cisplatin-type cross-linkers is their inherent DNA-targeted character. While intercalation plays an important role in the mechanism of action of these agents, rapid platination of DNA nucleobases appears to be crucial for potent inhibition of cancer cell proliferation. Characteristically, derivatives that react with DNA sluggishly or not at all have proven only marginally cytotoxic.^{16,17} Because the Pt–S_{cysteine} bond is resistant to nucleophilic attack by DNA nitrogen,² the GSH metabolites formed by compounds **1** and **2** (analogous to **I/I'** and **II/II'**, Scheme 1), even if they reach the nucleus, should be relatively nontoxic because of their inability to induce cytotoxic DNA adducts. Likewise, ACRAMTU (**III'**) and the bisintercalator **IV** (a reversible DNA binder^{18,19}), which are potential metabolites formed in the reaction between **2** and GSH, are considerably less cytotoxic than the hybrid agents.^{9,19}

Conclusions

In summary, we report the first case of a simple structural modification within a DNA-targeted platinum antitumor agent that increases the target affinity of the metal while concomitantly reducing unwanted reactivity with protein sulfur. On the basis of these findings, the amidine analogue, **1**, should have a major pharmacological advantage over PT-ACRAMTU (**2**), which may contribute to the superior potency of the new analogue *in vitro* and *in vivo*. To test this hypothesis, future studies will delineate relationships between the cytotoxicity and intracellular distribution of **1**, PT-ACRAMTU, and related complexes in cancer cells characterized by elevated cytosolic GSH levels. Another important issue to be addressed is the relatively high toxicity of our new hybrid agent in treated animals. The results of a necropsy performed on the test animals treated with sublethal doses of the hybrid agent (unreported data) revealed mild to yellow discoloration of the kidneys. We speculate that the

adverse effects on the kidneys are the result of platinum binding to sulfur of glutathione (GSH). Cisplatin–GSH adducts are metabolized to strong nephrotoxins that cause damage to the renal proximal tubules.² Thus, additional structural modifications in compound **2** may be needed to reduce this toxicity to levels as low as those observed for picoplatin and carboplatin.

Experimental Section

Materials. The platinum–acridine complexes **1** and **2** were synthesized according to published procedures.^{8,9} For the preparation of biological buffers, biochemical grade chemicals (Fisher/Acros) were used. HPLC grade solvent was used in all chromatographic separations. All other chemicals and reagents were purchased from common vendors and used without further purification. Stock solutions of the platinum compounds were prepared immediately before use.

NMR Spectroscopy. NMR spectra in arrayed experiments were collected at room temperature on a Bruker 500 DRX spectrometer equipped with a triple-resonance broadband inverse probe and a variable temperature unit. Reactions were performed in 5 mm NMR tubes containing 2 mM complex **1** or **2** and 2 mM *N*-acetylcysteine (10 mM phosphate buffer, D₂O, pH* 6.8). The 1-D ¹H kinetics experiments were carried out as a standard arrayed 2-D experiments using a variable-delay list. Incremented 1-D spectra were processed exactly the same, and suitable signals were integrated. Data were processed with XWINNMR 3.6 (Bruker, Ettlingen, Germany). The concentrations of platinum complex at each time point were deduced from relative peak intensities, averaged over multiple signals to account for differences in proton relaxation.

Incubations and Chromatographic Separations. Reactions of **1** and **2** with *N*-acetylcysteine were performed at a 1:1 drug-to-amino acid ratio in 10 mM sodium phosphate buffer (pH 7.1). Incubations were performed at 37 °C for 6 h. The mixtures were separated by reverse-phase HPLC using the LC module of an Agilent Technologies 1100 LC/MSD trap system equipped with a multiwavelength diode-array detector and an autosampler. A 4.6 mm × 150 mm reverse-phase Agilent ZORBAX SB-C18 (5 μm) analytical column was used in all of the assays, which was maintained at 25 °C during separations. A monitoring wavelength of 413 nm was used to detect acridine-containing fragments. The following eluent systems were used for the separations: solvent A, degassed water/0.1% formic acid, and solvent B, methanol/0.1% formic acid. The gradient used in the separation was 98% → 30% solvent A over 30 min at a flow rate of 0.5 mL/min. Peak integration was done using the LC/MSD Trap Control 4.0 data analysis software.

Mass Spectrometry. Mass spectra were recorded on an Agilent 1100LC/MSD ion trap mass spectrometer. After separation by *in-line* HPLC, adducts were directly infused into the atmospheric-pressure electrospray source. Ion evaporation was assisted by a flow of N₂ drying gas (350 °C) at a pressure of 40 psi and a flow rate of 10 L/min. Positive-ion mass spectra were recorded with a capillary voltage of 2800 V and a mass-to-charge scan range of 150 to 2200 *m/z*.

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Supporting Information Available: Plots of arrayed NMR spectra, HPLC profiles of the reaction mixtures, electrospray mass spectra, and tabulated mass data for all identified products. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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